

# International Journal of Recent Advances in Multidisciplinary Research

Vol. 02, Issue 12, pp.1110-1113, December, 2015

# **RESEARCH ARTICLE**

# LATENT EFFECT OF CLEOMEDROSERIFOLIA ETHANOLEXTRACT ON THE DEVELOPMENT OFOCHLEROTATUSCASPIUS AND CULEXPIPIENS

# 1\*Omnia, M.H.M. Kamel and <sup>2</sup>Mosaad, M. Hassan

<sup>1</sup>Department of Applied Chemistry, National Research Centre, Dokki, Cairo, Egypt <sup>2</sup>Department of Entomology, Faculty of Science, Ain Shams University, Cairo, Egypt

# **ARTICLE INFO**

# Article History:

Received 28<sup>th</sup>, September 2015 Received in revised form 15<sup>th</sup>, October 2015 Accepted 16<sup>th</sup>, November 2015 Published online 30<sup>th</sup>, December 2015

### Keywords:

Cleomedroserifolia, Biological activity, Mosquitoes.

# **ABSTRACT**

Mosquitoes are the most important vectors for serious human and animal diseases causing millions of deaths every year. Among these diseases, malaria, yellow fever, dengue and dengue hemorrhagic fever, filariasis and Rift Valley fever at endemic and epidemic areas in many countries. The ethanolic extract of *Cleomedroserifolia* was evaluated against *Culexpipiens* and *Ochlerotatuscaspius* LC<sub>50</sub> were 178.61 & 162.63 respectively. The pupation percent was significantly reduces as a result of extract application *Culexpipiens*. The adult emergence was also reduced in *Ochlerotatuscaspius* more than *Culexpipiens*. The gonotrophic cycle was elongated to 8.53 and 7 for *Ochlerotatuscaspius* and *Culexpipiens* respectively. The fecundity and fertility was significantly reduced. *Cleomedroserifolia* extract can be used as larvicide and insect growth regulator agent in the controlling program for mosquitoes.

# **INTRODUCTION**

Mosquitoes are the most important vectors for serious human and animal diseases causing millions of deaths every year. Among these diseases, malaria, yellow fever, dengue and dengue hemorrhagic fever, filariasis and Rift Valley fever at endemic and epidemic areas in many countries. The transmission of diseases can be interrupted by controlling the vectors by different ways (WHO, 1991, Lerdthusnee, et al., 1995 and Madani, et al., 2003). The continuous using of traditional chemical insecticides leads to adverse effects such as developing of resistance, affect the public healthand reducing beneficial non-target biota. Retardation of these effects forced authors to search for new products that are environmentally safe (Abbassy, 1998; Mohamed et al., 2003, Kamel et al., 2005 and Bakr et al., 2008). Cleome species leaf extract was evaluated against Callosobruchusmaculatus show complete inhibition of embryonic development of eggs exposed for 48h in all botanical treatments (Dabire, 2008). Different fraction extracted from Thymus vulgaris were tested for mosquitocidal activity against Culexquinquefasciatus. The oils showed highly efficiency with respect to mortality and to the percentage of adult emergence upon short-term exposure in water treated by lethal doses of individual oils. Furthermore, mortality increased significantly in relation to time of exposure, and total mortality of the larvae at the end of their development was about 90%. Such significant mortality was also naturally reflected in the total emergence of adultsPavela (2009).

\*Corresponding author: Omnia, M.H.M. Kamel,

Department of Applied Chemistry, National Research Centre, Dokki, Cairo, Egypt.

The authors do a continuous effort to improve the potency of plant extract as insecticide and using them alternate the traditional insecticides in IVM program.

# **MATERIALS AND METHODS**

# Preparation of plant extract

The tested plant (*Cleomedroserifolia*) was washed toavoid dusts and dirt then lift to dry undershade in the laboratory. Dried part of plant was cut into small pieces and ground in anelectric grinder. Hundred grams of theresulting powdered was exhaustively extracted with ethanolabsolute, following the method described by Kamel *et al.*, (2005).

# **Tested insects**

Culexpipiens and Ochlerotatus caspius (Culicidae: Diptera). Both species were provided from Medical Entomology Institute and transferred to the laboratory of Entomology Department – Faculty of Science – Ain Shams University where self-perpetuating colonies were established and maintained during the present study. The maintaining of colonies in laboratory was proceeded according to methods described by (Kamel et al. 2005 and Shaalan, et al. 2005).

# Larvicidal bioassay

The bioassay experiments were carried out according to (Mallick, et al. 2015). The investigation effect of sub-lethal concentration on the pupation, adult emergence and reproductive potential of both studied species were carried out as follow:

The 3<sup>rd</sup> instar larvae (500 larvae) were exposed to sub lethal dose of Cleomedroserifolia extract and Esbiothrin. Survived larvae after twenty-four hours post treatment were gently washed and transferred to labeled pans. Treated larvae and controls were kept under laboratory conditions of 27±2 °C and 75±5 % RH and a photoperiod of 16-8 hrs (L-D). Pupae were collected and placed in breeding cages. Emerged adults were countedand provided with cotton pads soaked with sugar solution (10%). After 2 days provide blood meal to Culex sp. Then each female was transferred individually into propylene tubes (4cm in diameter and 7cm depth) one third of which was filled with water and covered with muslin. Adults were observed daily for egg laying. Calculate the duration of the gonotrophic cycle, the females were observed daily for oviposition. The number of eggs laid by each female was counted to calculate the fecundity. The female fertility was calculated as the mean number of hatched eggs / the oviposited eggs per female.

# Statistical analysis

Calculate the LC<sub>50</sub> and LC<sub>95</sub> values through Log Probit analysis (Finney, 1971). Use Microsoft excel program (Mean  $\pm$  SD & Z-test).

# **RESULTS**

Ethanolic extract of *Cleomedroserifolia* showed larval mortality against both species used in present study. The  $LC_{50}$  and  $LC_{95}$  for *Culexpipiens* and *Ochlerotatuscaspius*were represented in table (1) and regression line in (Fig. 1). The result showed that the mortality increased with increasing the concentration of extract. *Ochlerotatuscaspius* is more susceptible to the extract than *Culexpipiens* larvae according to their  $LC_{50}$  162.63 and 178.61 ppm respectively.

Table 1. Larval mortality and LC<sub>50</sub> & LC<sub>95</sub> values for the *Cluexpipiens* and *Ochlerotatuscaspius* larvae exposed to *Cleome dorserifolia* ethanolic extract

Mosquito species	Conc.	Mortality %	LC50	LC95	Slope
	ppm		(Coefficient limit)	(Coefficient limit)	
Culexpipiens	80	13.3	178.61	666.13	2.9
	140	41.7	(157.56 - 202.45)	(485.53 - 915.17)	
	200	58.3			
	300	70			
	400	85			
Ochlerotatuscaspi	80	16.7	162.63	540.87	3.2
us	140	43.3	(144.28 - 183.29)	(415.73 - 704.42)	
	200	61.7			
	300	73.3			
	400	93.3			

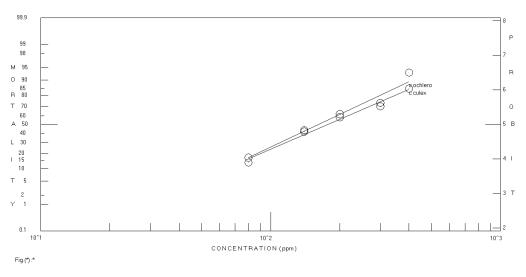


Fig. 1. Susceptibility of Culexpipiens and Ochlerotatuscaspius larvae to ethanolic extract of Cleome dorserifolia

Table 2. Efficiency of Cleomedorserifolia on some biological activities of autogenous Ochlerotatuscaspius and Culexpipiens

Contribution	Control	Ochlerotatuscaspius	Control	Culexpipiens
Survived larvae	500	75 (375)	500	75.4 (377)
% (n)				
Pupation	95.2 (476)	84.27 (316)	97 (485)	51.7 (195)
% (n)		* 2.96		*1.00
Emergence	98.7 (470)	49.37 (156)	99.2 (481)	73.8 (144)
% (n)		* 10.21		*1.00
Gonotrophic cycle	$3.47\pm0.52+$	$8.53 \pm 0.52^{+}$	$3.00 \pm .000 +$	$7 \pm 0.89^{+}$
Fecundity	$97.533 \pm 3.98 +$	$30.40 \pm 1.92^{+}$	$100.80 \pm 4.296 +$	$54.1 \pm 5.68^{+}$
Hatchability	$85.40 \pm 5.01 +$	$6.53 \pm 1.19^{+}$	$89.33 \pm 5.354 +$	$17.5 \pm 2.73^{+}$
(%)		(21.48)		(32.3)
		*15.31		* 1.00

The sub-lethal concentration of ethanolic extract of Cleomedorserifolia affect the life cycle of both species from mosquitoes. The results represented in Table (2) showed different variation in life cycle during pupation period and adult emergence also gonotrophic cycle, fecundity and hatchability. There was significantly reduction in pupation for Culexpipiens but in Ochlerotatuscaspius the reduction during adult emergence. In both species fecundity and hatchability were significantly reduction. Also, the gonotrophic cycle was elongate comparing with the control of both species (autogenus and anatogenus, Ochlerotatuscaspius and Culexpipiensrespectively.

# **DISCUSSION**

Mosquitoes are still the world wide vector of human and animal diseases, and are conspicuous nuisance pests as well, even after massive efforts of control. The extensive uses of chemical insecticides have induced resistance by insect pests, caused contamination of human food, mammalian toxicity, reducing beneficial non-target biota and environmental pollution. Active substances extracted from plants are used as insect repellents or synergists (Thangam and Kathiresan, 1997; El-Gougary, 1998 and Mansour et al., 2000), or effective as insect growth regulator and adulticide (Beehler and Mulla, 1993; Mc Carry, 1996, Kassem et al., 2000 and Nathan et al. 2006). Various parts of the plants have been reported by researchers which give insecticidal activity against different species of mosquitoes (Hossain et al., 2011; Singha and Chandra, 2011; Adhikari et al., 2012; Mallick et al., 2014; Singha Ray et al., 2014 and Singha et al., 2015).

The tested extract revealed differences in LC<sub>50</sub>, LC<sub>95</sub> values and the slope functions of the regression lines. The Ochlerotatuscaspius is more susceptible to the extract than Culexpipien. This result may be attributed to the reaction of each species toward the same plant extract. Mallick et al., 2015 reported that different species of mosquitoes appear different degree of susceptibility toward the ethanolic leaf extract of Annonareticulate. The sub-lethal concentration of ethanolic extract of Cleomedroserifolia showed significant reduction in some biological activities of both species of mosquitoes. The treatment with extract revealed significantly reduces in pupation percent for Culexpipiens may be attributed to the high tolerance of Ochlerotatuscaspius than Culexpipiens during pupal stage. Reduction in the adult emergence of different mosquito species treated with different plant extracts and pyrethroids was previously recorded by several investigators (Pereira and Gurudutt, 1990; Mohsen et al., 1990; Awala et al., 1998; Pushpalatha and Muthukrishnan, 1999; Attiaa, 2002; EL-Bokl, 2003 and Abd El-Bakyet al, 2005). Also, Shaalan et al., 2005 reported that the duration of developmental stages of Aedesaegypti were prolonged when treated with synthetic and botanical insecticides.

In present study, the adult emergence was affected also by treatment and gonotrophic cycle was elongated. The length of the gonotrophic cycle is of crucial importance in determining the vectorial capacity of the mosquitoes in disease transmission. However, the effect of larvicidal treatment on the avidity of the emerged females to acquire blood meals needed to be investigated. Marcardet al., (1986) had indicated that the females of Ae. aegypti, Ae. togoi and Cx.

quinquefasciatus developed from treated larvae with methanol extracts of Ajuga spp. took smaller blood-meals than those developing from untreated ones.

The reduction of fecundity may be due to lack of proteins required for the formation of yolk granules, small blood meal intake or inhibition of protein synthesis due to the treatment may lead to the reduction the number of deposited eggs. and fertility were significantly reducesmay correlate either from deficiency in the egg protein or the reduction of the sperms viability and the probable presence of sperm abnormalities. These reasons may affect the embryonic development which may lead to reduction of egg hatchability. All these disorder may correlate with some biochemicalchanges in the tested species which may lead to certain functional and physiological interactions (Teller, 1965; El-Bokl *et al.*,1998; Mohamed and Hafez, 2000; Massoud *et al.*, 2001; Abd El-Baky *et al.*, 2005; Helmy *et al.*, 2010 and Bakr *et al.*, 2010).

# Conclusion

From the present study the author can concluded that, the *Cleomedroserifolia* exthanolic extract can be used as larvicide or insect growth regulatorfor mosquito species.

# REFERENCE

Abbassy, M.A., El\_Gougary, O.A., El\_Hamady, S., Sholo, M.A. 1998. Insecticidal, acaricidal and synergistic effects of soosan, Pancratiummaritimum extracts and constituents. *J. Egypt. Soc. Parasitol.* 28(1):197-205.

Abd El-Baky, S.M.M., Hafez, J.A., Hamed, M.S., Kamel, O.M. 2005. Latent effect of Cleome droserifollia extract, Esbiothrin and Alsystin treatments on some biological activities of Ochlerotatuscaspius (Diptera: Culicidae). Egypt. Acad. Soc. Environ. Develop. (A-Entomology) 6 (3): 51-60.

Adhikari, U., Singha, S., Chandra, G. 2012. In vitro repellency and larvicidal efficacy of *Swieteniamahagoni* against the larval forms of *Culexquinquefasciatus* Say. *Asian Pac. J. Trop. Biomed.* S260 – S264.

Attiaa, S.R.A. 2002. Ph.D. Thesis Ent. Dept. Ain shams Univ. Cairo, Egypt.

Awala, P.,Mawangi, R.W., Irungu, L.W. 1998. Larvicidal activity of a granular formulation of a Meliavolkensii (Gurke) acetone extract against Ae. aegypti L.. Insect Science and its Application. 18 (3): 225-228.

Bakr, R.F.A., Helmy, N.A., Nawwar, G.A., Ibrahim, S., El, Helmy, O.M. 2010. Changes in protein content of *Culexpipiens* mosquito treated with two agriculture waste extracts. *J. Egypt Acad. Biolog. Sci.* 3 (1): 95-103.

Bakr, R.F.A., Hussein, M.A., Hamouda, L.S., Hassan, H.A., Elsokary, Z.F. 2008. Effect of some insecticidal agents on some biological aspects and protein patterns of desert locust Schistocercagregaria (Forskal). *Egypt. Acad. Soc. Environ*. Develop. 9 (2): 29-42.

Bhatnagar, M., Kapur, K.K., Jalees, S., Sharma, S.K., 1993. Laboratory evalution of insecticidal properties of Ocimumbasilicum Linnaeus and O. sanctum Linnaeus plant's essential oils and their major constituents against vector mosquito species. *J. Entomol. Res.* 17 (1): 21-26.

Dabire, C.L.B., Niango, Ba, M., Sanon, A. 2008. Effects of crushed fresh Cleome viscosa L. (Capparaceae) plants on

- the cowpea storage pest, Callosobruchusmaculatus Fab (Coleoptera: Bruchidae). *Int. J. Pest Manag.* 54 (4): 319 326.
- EL-Bokl, M.M. 2003. Latent toxicity of Azadirachtin treatment on Culexpipiens (Diptera: Culicidae). J. Egypt. Acad. Soc. Environ. Develop. (A- Ent.) 3 (1): 63-74.
- El-Bokl, M.M., Hamed, M.S., Moawad, H.M. 1998. Effects of sublethal dosage of plant extracts and Cypermethrin on certain biochemical aspects of the first of filial generation of Culexpipiens larvae. *J. Egypt. Ger. Soc. Zool.* 26 (E): Entomology.
- El-Gougary, O.A. 1998. Insecticidal and synergistic activity of Atriplexhalimus L. extracts. *J. Egypt Soc. Parasitol.* 28 (1): 191 196.
- Finney, D.J. 1971. Probit Analysis (3rd ed.). Cambridge Univ. Press, 333 p. Cambridge.
- Helmy, N.A., Bakr, R.F.A, Nawwar, G.A., Ibrahim, S.E.I, Helmy, O.M. 2010. Biochemical effects of some agricultural waste extracts against *Culexpipiens* (Diptera: culicidae). *J. Egypt Acad. Biolog. Sci.* 2(2): 75-81.
- Hossain, E., Rawan, A., Chandara, G., Mandal, S.C., Gupta, J.K. 2011. Larvicidal activity of *Dregeavolubilis* and *Bombaxmalabaricum* leaf extracts against the filarial vector Culexquinquefasciatus. *Asian Pac. J. Trop. Med.* 4: 436 – 441.
- Kamel, O. M., Hassan, M. M., Abd, El-Baky. S.M.M., Hafez, J.A., Hamed, M.S. 2005. Effect of the salinity and temperature on the larvicidal activity of some insecticidal agents the mosquito Ochlerotatuscaspius (Diptera: Culicidae). J. Egypt. Acad. Soc. Environ. Develop. (A. Entomology) 6 (2): 307-321.
- Kassem, H.A., Tewfick, M.K., El-Sawaf, B.M. 2000. Evaluation of avermectins as sand fly control. *J. Egypt Ger. Soc. Zool.* 31: 1 11.
- Lerdthusnee, K., Romoser, W.S., Faran, M.E., Dohm, D.J. 1995. Rift Valley Fever virus in the cardia of Culexpipiens an immunocytochemical and Ultrastructural study. *Am. J. Trop. Hyg.* 53 (4): 331 7.
- Madani, T.A., Al-Mozrou, Y.Y., Al-Jeffri, M.H., Mishkhas,
  A.A., Al-Rabeah, A.M., Turkistani, A.M., Al-Sayed, M.O.,
  Abodahish, A.A., Khan, A.S., Ksiazek, T.G., Shobokshi, O.
  2003. Rift valley fever epidemic in Saudi arabia: epidemiological, clinical and laboratory characteristics.
  Oxford Journal.Clinical Infectious Diseases. 37(8): 1084 1092.
- Mallick, S., Banerjee, R., Chandra, G. 2015. Mosquito larvicidal potential of ethanol leaf extract of the plant, Annona reticulate L. against Aedesaegypti L. and Culexquinquefasciatus Say (Diptera: Culicidae). *J. Mosq. Res.* 5 (19): 1 7.
- Mallick, S., Bhattacharya, K., Chandra, G. 2014. Mosquito larvicidal potentiality of wild turmeric *Curcumaaromatic* rhizome extracts against Japanese Encephalitis vector *Culexvishnui*group. *J. Mosq. Res.* 4 (19): 1 6.
- Mansour, S.A., Messeha, S.S., El-Gengaihi, S.E. 2000. Botanical biocides 4. Mosquicidal activity of certain Thymus capitatus. Constituents. *J. Nat. Toxins.* 9 (1): 49-62.
- Marcard, M., Zebitz, C.P.W., Schmutterer, H. 1986. The effect of crude methanolic extracts of Ajuga spp. on the developmental stages of various mosquito species. *J. Appl. Entomol* 101 (2): 146-154.

- Massoud, A.M., Labib, I.M., Rady, M. 2001. Biochemical changes of Culexpipiens larvae treated with oil and oleo resin extracts of Myrrh commiphoramolmol. *J. Egypt. Soc. Parasitol.* 31 (2): 517-529.
- Mc Carry, M.J. 1996. Efficacy and persistence of Altosid pellets against Culex species in catch basins in Michigan. *J. Am. Mosq. Control Assoc.* 12 (1): 144 146.
- Mohamed, M.I., El-Mohamady, R.H., Mohamed, H.A. 2003. Larvicidal activity and biochemical effects of certain plant oil extracts against Culexpipiens larvae (Diptera: Culicidae). *J. Egypt. Acad. Soc. Environ. Develop.* (A. Ent.) 3 (1): 75-93.
- Mohamed, M.I., Hafez, S.E. 2000. Biological and biochemical effects of the non volatile plant oil (Jojoba) against Culexpipiens (Diptera: Culicidae). *J. Egypt. Ger. Soc. Zool.* 13 (A): 65-78.
- Mohsen, Z.H., Jawad, A.L.M., Al-Chalabi, B.M., Al-Naib, A. 1990. Biological activity of Callistemon lanceolatus against Culexquinquefasciatus. Fitoterapia 61 (3): 270-274.
- Nathan, S.S. 2006. Effects of Meliaazedarach on nutritional physiology and enzyme activities of the rice leaf folder Cnaphalocrocismedinalis (Guenee) (Lepidoptera: Pyralidae). *Pestic.Biochem. Physiol.* 84: 98-108.
- Pavela, R., 2009. Larvicidal property of essential oils against *Culexquiquefasciatus* Say (Diptera: Culicidae). *J. Ind. Crops & Prod.* 30 (2): 311 315.
- Pereira, J., Gurudutt, K.N. 1990. Growth inhibition of Muscadomestica L. and Culexquinquefasciatus (Say) by (-)-3-epicaryoptin isolated from leaves of Clerodendroninerme (Gaertn) (Verbenaceae). *J. Chem. Ecol.* 16 (7): 2297-2306.
- Pushpalatha, E., Muthukrishnan, J. 1999. Efficacy of two tropical plants extracts for the control of mosquitoes. *J. Appl. Entom.* 123 (6): 369-373.
- Shaalan, E.A., Canyon, P.V., Younes, M.W.F., Abdel-Wahab, H., Mansour, A. 2005. Effect of sub-lethal concentrations of synthetic insecticides and *Callitrisglaucophylla* extracts on the development of *Aedesaegypti*, *J. Vec.Eco.* 30 (2): 295 298.
- Singh, A., Bhattacharya, K., Chandra, G. 2015. Efficacy of *Nicotianaplumbaginifolia* (Solanaceae) leaf extracts as larvicide against malarial vector Anophielesstephensi (Liston 1901). *Int. J. Pharma. Bio. Sci.* 6 (1): (B): 860 868
- Singha Ray, A., Bhattacharya, K., Singh, A., Chandra, G. 2014. Larvicidal activity of *Nelumbonucifera*Gaertn (Nymphaeaceae) against Anopheles stephensi (Liston 1901) and its effect on non-target organism. *J. Mosq. Res.* 4 (10): 1 7.
- Singha, S., Chandra, G. 2011. Mosquito larvicidal activity of some common species and vegitable waste on *Culexquinquefasciatus* and *Anophelesstephensi*. Asian Pac. *J. Trop. Med 4* (4): 288 93.
- Teller, W.H. 1965. The mechanism and control of yolk formation. Ann. Rev. Entomol., 161-184.
- Thangam, T.S., Kathiresan, K. 1997. Mosquito larvicidal activity of mangrove plant extracts and synergistic activity of Rhizophoraapiculata with pyrethrum against Cx. quinquefasciatus. *Inter. J. Pharmacognosy*, 69 71.
- World Health Organization (WHO) 1991. specialprogramme for research and training in tropical diseases (TDR). Geneva. Swizerland.